

## Sydnonimines as exogenous NO donors

*E. Yu. Khmel'nitskaya,<sup>a</sup> V. I. Levina,<sup>a</sup> L. A. Trukhacheva,<sup>a</sup> N. B. Grigoriev,<sup>a</sup> V. N. Kalinin,<sup>b</sup>  
I. A. Cherepanov,<sup>b</sup> S. N. Lebedev,<sup>b</sup> and V. G. Granik<sup>a\*</sup>*

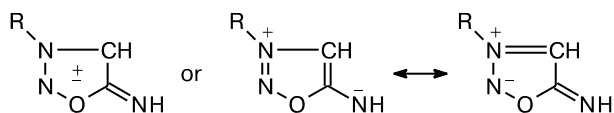
<sup>a</sup>Antibiotic State Scientific Center,  
3a ul. Nagatinskaya, 117003 Moscow, Russian Federation  
Fax: +7 (095) 231 4284. E-mail: vggranik@mail.ru.

<sup>b</sup>A. N. Nesmeyanov Institute of Organoelement Compounds, Russian Academy of Sciences,  
28 ul. Vavilova, 119991 Moscow, Russian Federation.  
Fax: +7 (095) 135 5085. E-mail: vkalin@ineos.ac.ru

Chemical oxidation of a series of sydnimine derivatives followed by NO release was studied. Substances having alkylamine substituents in the position 3 were shown to be considerably more potent NO donors in comparison with those having alkyl or aralkyl substituents in the position 3. It was suggested that the effect is mainly due to lowering of the activation energy of NO release upon stabilization of the cation formed competitively by the amino group.

**Key words:** sydnonimines, 3-aminosydnonimines, nitrogen oxide, NO donors.

Sydnonimines represent a class of mesoionic heterocyclic compounds exhibiting a broad range of biological activities and capable of functioning as nitrogen oxide donors<sup>1–3</sup> (a review on the synthesis and properties of sydnonimines has been reported<sup>4</sup>).

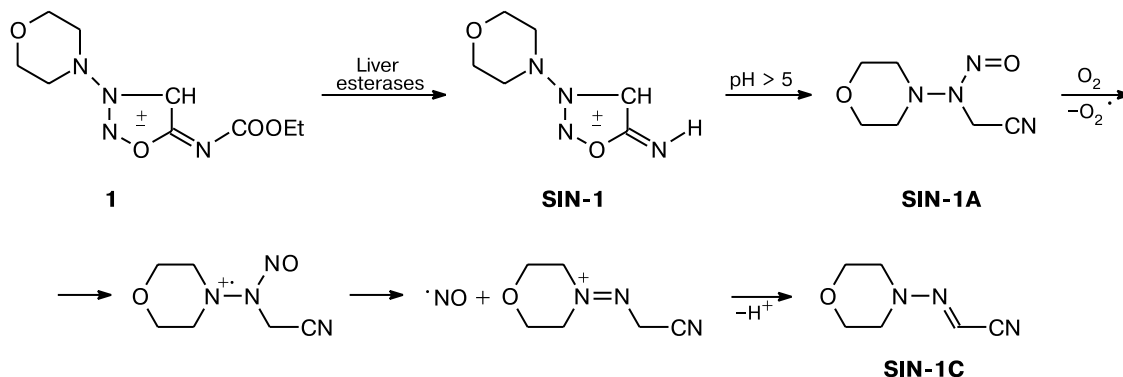


For example, it has been found that the biological action of the known antianginal drug molsidomine (**1**) is related to its ability to release nitrogen oxide (Scheme 1).

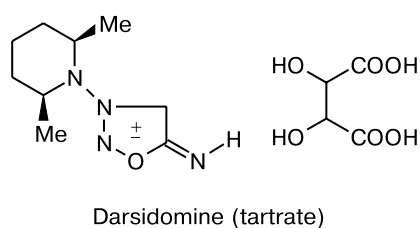
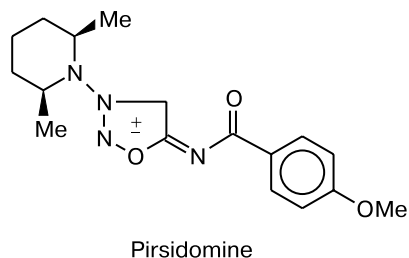
Decomposition of **SIN-1A** (unlike that of **SIN-1**) is a pH-independent process and involves oxygen.<sup>2</sup> Oxygen consumption in buffered solutions of **SIN-1** is correlated

with the formation of nitrogen oxide. It is significant that NO rather than HNO is formed from **SIN-1A** together with **SIN-1C**. The compound **SIN-1A** is the activator of soluble guanylate cyclase (sGC); it increases the intracellular concentration of cyclic guanosine monophosphate (cGMP) and, correspondingly, enhances its effects (vasodilation, inhibition of thrombocyte aggregation, neurotransmission, immune response regulation) typical of nitrogen oxide donors.<sup>5</sup> The drug can undergo either enzymatic or nonenzymatic degradation; under physiological or more alkaline conditions, **SIN-1** undergoes fast nonenzymatic ring cleavage to give **SIN-1A**, which is quite stable at pH 7.4 under anaerobic conditions in solutions protected from light. However, even traces of oxygen promote further transformations up to the formation of *N*-morpholinoiminoacetonitrile (**SIN-1C**).<sup>6</sup>

Scheme 1



Worth noting are several other compounds of the sydnonimine series that attract considerable attention due to the high biological activity. These are molsidomine analogs such as pirsidomine, which is a potent vasodilator with prolonged action.<sup>7–9</sup> Like molsidomine, pirsidomine is first metabolized in the organism, and the metabolites function as nitrogen oxide donors.<sup>7</sup> The major pirsidomine metabolite is darsidomine (used as the tartrate), which causes selective dilatation of the coronary artery and has an anti-ischemic action without inducing tolerance on long-term administration.<sup>10–12</sup>



Similarly to sydnonimines, derivatives of other meso-ionic compounds also exhibit clear-cut nitrogen oxide donor properties and the corresponding biological effects. Solutions with pH 6.2–6.8 were shown<sup>13</sup> to be the optimal media for nitrogen oxide release from mesoionic compounds of the oxatriazolium structure. The mechanism of nitrogen oxide release proposed in the study cited<sup>13</sup> differs somewhat from that presented above. It was specially noted that the pathways of the *in vitro* and *in vivo* NO generation do not coincide; in the latter case, the release of nitrogen oxide can be enhanced due to the enzymatic degradation in the presence of thiols. Note that an alternative degradation route giving rise to the nitroxyl, HNO, instead of nitrogen oxide was also indicated for these compounds. It is also noteworthy that some known drugs, namely, the psychostimulant sydnocarb and the mono-aminoxidase inhibitor sydnophen have also proved to be fairly active nitrogen oxide generators.<sup>14,15</sup>

Although a large number of publications have been devoted to the properties of sydnonimines,<sup>15</sup> the relationship between the NO-donor properties and the structures of these compounds has not been adequately elucidated.

The present work continues our research into the effect of the structures of sydnonimines on their chemical, hydrolytic, oxidative and, especially, NO-donor properties depending on the position and the nature of substitu-

**Table 1.** Yield of nitrogen oxide on chemical oxidation\* of sydnonimine derivatives

Compound	Structure	Yield of NO (%)
1		14±2
2		31±2
3		19±2
4		22±2
5		22±2
6		1±0.2
7		0.7±0.2
8		0.7±0.2
9		3.6±0.2

\* Oxidation conditions: 0.1 M NaOH, 80 °C.

ents in the sydnonimine ring. This may provide the base for the search for new biologically active compounds in the sydnonimine series.

The structures of compounds studied in this work are summarized in Table 1. As can be seen by examining their formulas, these compounds can be divided into two large groups: substances bearing amine-containing substituents (1–5) in position 3 of the sydnonimine ring, and those having alkyl substituents in this position (6–9). In both groups, some compounds contain acyl residues at the endocyclic nitrogen atom (1, 3–5 in the first group and 8, 9 in the second group). In addition, the first group comprises compounds with arylalkyl substituents in position 4 (4, 5), while compounds 8 and 9 belonging to the second group have a phenyl substituent at this position.

The properties of the sydnonimines as nitrogen oxide donors were compared in this study with the properties of molsidomine (1) and sydnophen (6). The substituted sydnonimines 2–5 and 7–9 were synthesized from the basic compounds of this series such as 2, 3, and 3-methyl-



formation considered above and the known<sup>17</sup> ring transformation of sydnonimines into oxotriazoles.

It is noteworthy that the results obtained in this study are in good agreement with the tentative results obtained in the study of activation of the enzyme sGC by these compounds (the biochemical studies were carried out by prof. I. S. Severina at the Research Institute of the Biomedical Chemistry of the Russian Academy of Medical Sciences, the results will be published elsewhere). Soluble guanylate cyclase (sGC) catalyzes the biosynthesis of so-called secondary messenger, *i.e.*, cyclic guanosine monophosphate from guanosine triphosphate.

Study of various preparations on sGC is a convenient and illustrative test to verify the ability of substances to release nitrogen oxide, as the non-heme mechanism of sGC activation is exceptionally seldom found. In our case, sGC activation by first-group compounds exceeds appreciably the activation caused by the second-group sydnonimines.

The results obtained imply that the nitrogen oxide donors most efficient in this series should be selected among compounds containing amino substituents in position 3 of the sydnonimine ring, which may prove to be a rational approach to the search for new biologically active compounds.

### Experimental

<sup>1</sup>H NMR spectra (DMSO-*d*<sub>6</sub>) were recorded on a Bruker WM-400 spectrometer. Melting points were determined in a glass capillary in a metallic block and were not corrected. All reactions involving organometallic compounds were carried out under dry argon in anhydrous solvents. The absorbance of solutions with an azo dye were measured on a KFK-3-01 photometer at a wavelength of 542 nm.

3-Methyl-4-phenylsydnonimine hydrochloride (**10**),<sup>18</sup> 3-butylsydnonimine hydrochloride (**7**),<sup>19</sup> 3-dimethylaminosydnonimine hydrochloride (**2**),<sup>20</sup> and 3-dimethylamino-*N*(6)-acetylsydnonimine (**3**)<sup>20</sup> were synthesized by previously described procedures.

**3-Methyl-4-phenyl-*N*(6)-pivaloylsydnonimine (8).** Pivaloyl chloride (2.46 mL, 2.4 g, 0.02 mol) was added to a suspension of compound **10** (2.12 g, 0.01 mol) in 10 mL of THF and the mixture was cooled to 0 °C. Pyridine (4.04 mL, 3.95 g, 0.05 mol) was added dropwise. The reaction mixture was stirred for 5 h at 0 °C and allowed to stand for ~14 h. The solvent was evaporated *in vacuo* and the residue was treated with 20 mL of water and 30 mL of chloroform. The layers were separated, the aqueous layer was extracted with chloroform (2×15 mL), and the combined organic extracts were dried with sodium sulfate and concentrated. The residue was recrystallized from ethanol to give 2.33 g (90%) of compound **8**, m.p. 149–150 °C. Found (%): C, 64.55; H, 6.81; N, 15.90. C<sub>14</sub>H<sub>17</sub>N<sub>3</sub>O<sub>2</sub>. Calculated (%): C, 64.85; H, 6.61; N, 16.20. <sup>1</sup>H NMR, δ: 1.26 (s, 9 H, Bu<sup>t</sup>); 4.23 (s, 3 H, Me); 7.50–7.60 (m, 3 H, *p*- and *m*-H, Ph); 7.66–7.72 (m, 2 H, *o*-H, Ph).

**3-Methyl-4-phenyl-*N*(6)-trifluoroacetylsydnonimines (9).** Trifluoroacetic anhydride (2.8 mL, 4.2 g, 0.02 mol) was added

to a suspension of 3-methyl-4-phenylsydnonimine (2.12 g, 0.01 mol) in 10 mL of THF and cooled to 0 °C. Pyridine (2.73 mL, 2.0 g, 0.02 mol) was added dropwise to the resulting mixture. The mixture was stirred for 5 h at 0 °C and allowed to stand for ~14 h. The solvent was evaporated *in vacuo*, the residue was treated with 20 mL of water and 30 mL of chloroform. The layers were separated, the aqueous layer was extracted with chloroform (2×15 mL), and the combined organic extracts were dried with sodium sulfate and concentrated. The residue was recrystallized from ethanol to give 2.30 g (85%) of compound **9**, m.p. 183–184 °C. Found (%): C, 48.93; H, 2.55; N, 15.00. C<sub>11</sub>H<sub>8</sub>F<sub>3</sub>N<sub>3</sub>O<sub>2</sub>. Calculated (%): C, 48.72; H, 2.97; N, 15.49. <sup>1</sup>H NMR, δ: 4.35 (s, 3 H, Me); 7.57–7.69 (m, 5 H, Ph).

**4-Hydroxy-3-dimethylamino-(4-dimethylaminophenyl)methyl-*N*(6)-acetylsydnonimines (4).** A 1.6 *M* solution of Bu<sup>n</sup>Li in hexane (1.84 mL, 2.95 mmol) was added to a solution of compound **3** (0.5 g, 2.95 mmol) in 20 mL of THF at –78 °C. The mixture was stirred for 30 min and 4-dimethylaminobenzaldehyde (0.57 g, 3.82 mmol) was added to the resulting red-brown solution. The mixture was stirred for 15 min at –78 °C and cooling was terminated. After the reaction mixture warmed up to room temperature, water (1 mL) was added. The solvent was removed *in vacuo* and the residue was dissolved in chloroform and filtered through an Al<sub>2</sub>O<sub>3</sub> layer (2×3 cm), the product being eluted with chloroform. Chloroform was evaporated *in vacuo* and the residue was chromatographed on a SiO<sub>2</sub> plate (elution with a chloroform–ether mixture (5 : 1)). Recrystallization from an ether–hexane mixture (1 : 2) gave 0.66 g (70%) of compound **4**, m.p. 123–124 °C. Found (%): C, 56.22; H, 6.40; N, 21.50. C<sub>15</sub>H<sub>21</sub>N<sub>5</sub>O<sub>3</sub>. Calculated (%): C, 56.41; H, 6.63; N, 21.93. <sup>1</sup>H NMR, δ: 1.86 (s, 3 H, Me); 2.92 (s, 6 H, C<sub>6</sub>H<sub>4</sub>–N(CH<sub>3</sub>)<sub>2</sub>); 3.16 (s, 6 H, 4-NMe<sub>2</sub>); 5.93 (s, 1 H, CH–OH); 6.50–6.60 (m, 1 H, CH–OH); 6.67, 7.10 (both m, 2 H each, C<sub>6</sub>H<sub>4</sub>).

**4-Hydroxy-3-dimethylamino-(4-trifluoromethylphenyl)methyl-*N*(6)-acetylsydnonimine (5).** A 1.6 *M* solution of Bu<sup>n</sup>Li in hexane (1.84 mL, 2.95 mmol) was added to a solution of compound **3** (0.5 g, 2.95 mmol) in 20 mL of THF at –78 °C. The mixture was stirred for 30 min and 4-trifluoromethylbenzaldehyde (0.57 g, 3.82 mmol) was added to the resulting red-brown solution. The mixture was stirred for 15 min at –78 °C and cooling was terminated. After the reaction mixture warmed up to room temperature, water (1 mL) was added. The solvent was removed *in vacuo* and the residue was dissolved in chloroform and the solution was filtered through an Al<sub>2</sub>O<sub>3</sub> layer (2×3 cm), the product being eluted with chloroform. Chloroform was evaporated *in vacuo* and the residue was chromatographed on a SiO<sub>2</sub> plate (elution with a chloroform–ether mixture (5 : 1)). Recrystallization from an ether–hexane mixture (1 : 2) gave 0.76 g (75%) of compound **5**, m.p. 96–97 °C. Found (%): C, 48.91; H, 4.52; N, 16.33. C<sub>14</sub>H<sub>15</sub>N<sub>4</sub>F<sub>3</sub>O<sub>3</sub>. Calculated (%): C, 48.84; H, 4.39; N, 16.27. <sup>1</sup>H NMR, δ: 1.80 (s, 3 H, Me); 3.31 (s, 6 H, NMe<sub>2</sub>); 6.13 (s, 1 H, CH–OH); 7.43, 7.70 (both m, 2 H each, C<sub>6</sub>H<sub>4</sub>).

### References

1. A. Dendorfer, *HERZ*, 1996, **21** (Suppl. 1), 38.
2. K. Schonafinger, *II Farmaco*, 1999, **54**, 316.
3. C. G. Newton and C. A. Ramsden, *Tetrahedron*, 1982, **38**, 2965.

4. V. G. Yashunskii and L. E. Kholodov, *Usp. Khim.*, 1980, **49**, 54 [*Russ. Chem. Rev.*, 1980, **49** (Engl. Transl.)].
5. H. Kankaanranta, R. G. Knowles, P. Vuorinen, O. Kosonen, P. Holm, and E. Moilonen, *Molec. Pharmacol.*, 1997, **51**, 882.
6. H. Bohn and K. J. Schonafinger, *Cardiovasc. Pharmacol.*, 1989, **14** (Suppl. 11), S 6.
7. H. Bohn, R. Beyerle, P. A. Martorana, and K. J. Schonafinger, *Cardiovasc. Pharmacol.*, 1991, **18**, 522.
8. M. Feelisch, *Naunyn-Schmiedebergs Arch. Pharmacol.*, 1998, **358**, 113.
9. I. L. Megson, *Drugs Future*, 2000, **25**, 701.
10. *11 Annual Drug Data Rep.*, Ed. J. R. Prous, 1995, **16** (3), 254.
11. J. Wang, G. Zhao, W. Shen, M. Ochoa, D. Moore, J. W. Hubbard, and T. H. Hintze, *J. Cardiovasc. Pharmacol.*, 1993, **22** (Suppl. 7), S 51.
12. A. Mulsch, M. Hecker, and P. I. Modvintcev, *Naunyn-Schmiedebergs Arch. Pharmacol.*, 1993, **347**, 92.
13. P. Holm, M.-K. T. Kankaanranta, and E. Moilanen, *Eur. J. Pharmacol.*, 1998, **346**, 97.
14. V. I. Levina, N. B. Grigor'ev, and V. G. Granik, *Khim. Geterotsikl. Soedin.*, 2004, 604 [*Chem. Heterocycl. Compd.*, 2004 (Engl. Transl.)].
15. V. G. Granik and N. B. Grigor'ev, *Izv. Akad. Nauk. Ser. Khim.*, 2002, 1268 [*Russ. Chem. Bull, Int. Ed.*, 2002, **51**, 1375].
16. R. E. Saltzman, *Anal. Chem.*, 1954, **26**, 1949.
17. H. U. Daeniker and J. Druey, *Helv. Chim. Acta*, 1962, **45**, 2426.
18. V. F. Vasil'eva and V. G. Yashunskii, *Khim. Nauka Prom.' [Chemical Science and Industry]*, 1959, 678 (in Russian).
19. V. G. Yashunskii, O. I. Samoilova, and L. E. Kholodov, *Zh. Org. Khim.*, 1964, **34**, 2050 [*J. Org. Chem. USSR*, 1964, **34** (Engl. Transl.)].
20. M. Gotz and K. Grozinger, *J. Heterocycl. Chem.*, 1970, **7**, 123.

*Received June 25, 2004;  
in revised form August 5, 2004*